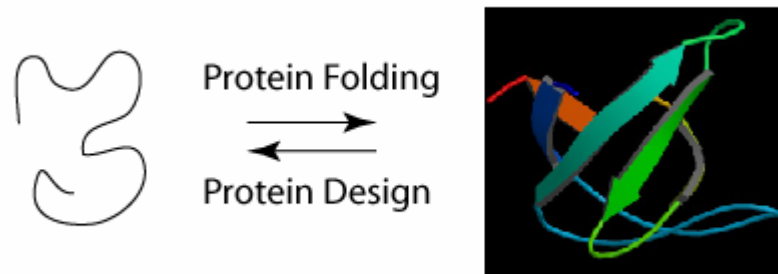


Protein design

- Branden & Tooze, Chapter 17
- Protein design is an extreme form of protein engineering, in which the entire polypeptide chain is built in its entirety rather than mutated piecemeal
 - may or may not involve designing the backbone
 - typically does not use an existing sequence
 - sometimes called “de novo” protein design to stress the “ground-up” approach
- Can be characterized as “inverse protein folding” because one starts from structure and predicts the amino acid sequence



- Large number of possible amino acid sequences makes the problem *NP*-hard
 - Pierce and Winfree, Protein Engineering 15, 779 (2001)

Knowledge-based design

Known structures contain information that may be used productively during a design study

statistical data, e.g. distribution of amino acids on a helix
modeling on a computer

Think small and dream big

introduce elements to form and stabilize secondary structures

hydrophobic patterns on a helix

engineer connectivity by introducing turns, loops,

Drawbacks

- The problem can quickly get out of hand due to the large number of variables that must be evaluated
- “Knowledge” is very personal and difficult to objectify



Helix bundles

Helix bundles are biologically important and common in many structural proteins, transcription factors (coiled coil), and enzymes

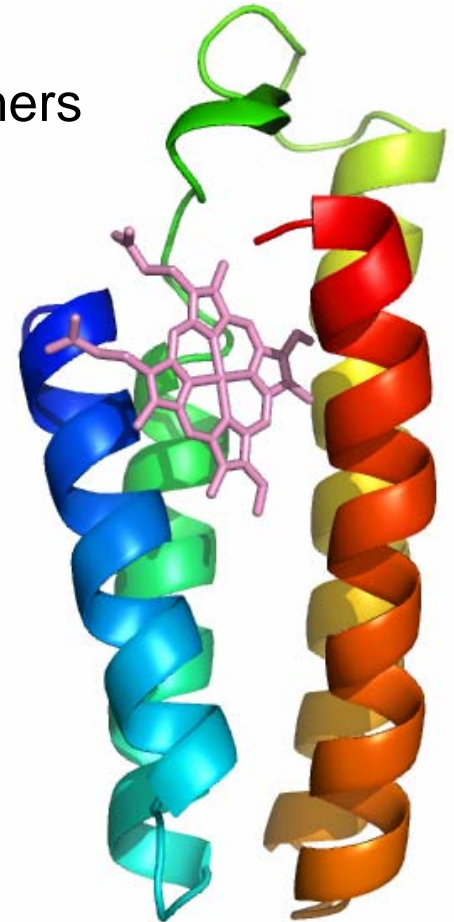
Stable and soluble—less likely to create run-away oligomers

Helices are easier to design de novo because they are stabilized through **local interactions**, and the factors that contribute to their formation are better understood

Easy to characterize experimentally

- CD, fluorescence

- thermal and chemical denaturation

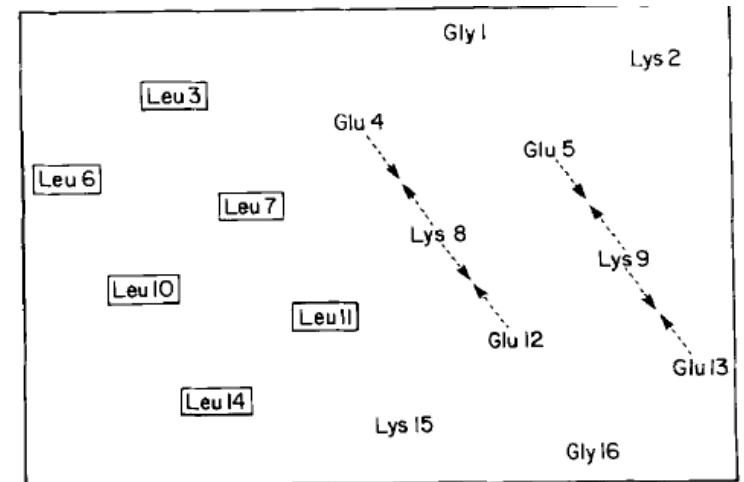
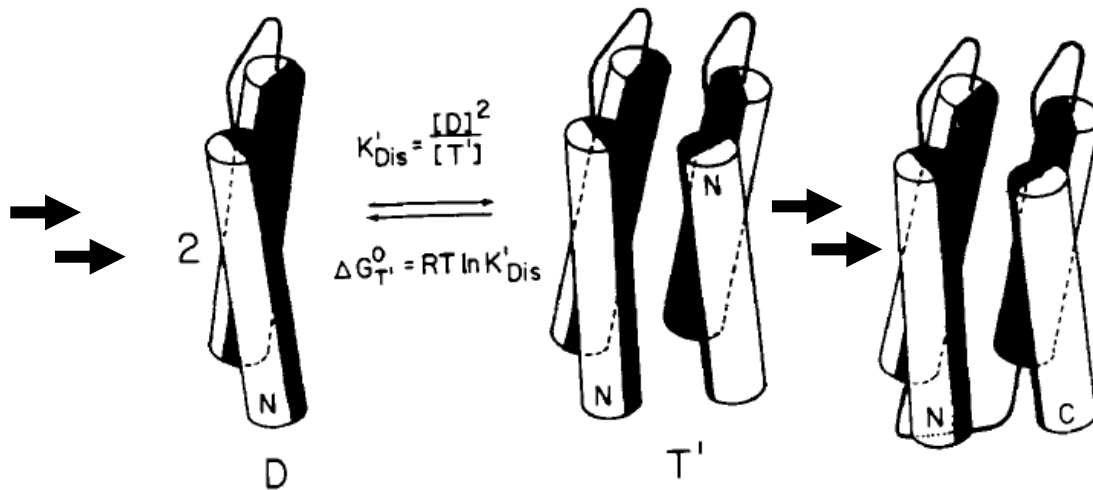
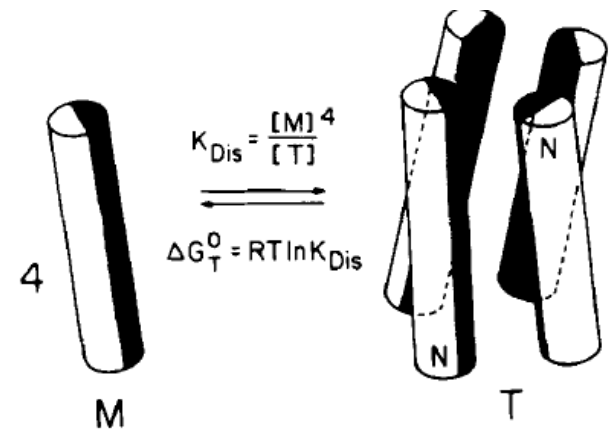


cytochrome b562

Designing helix bundles

Design 12 and 16 residue peptides that are likely to form helices and binary pattern them using Leu on one side, and Glu and Lys on the other on the other

Hydrophobic surfaces should come together to avoid contact with solvent

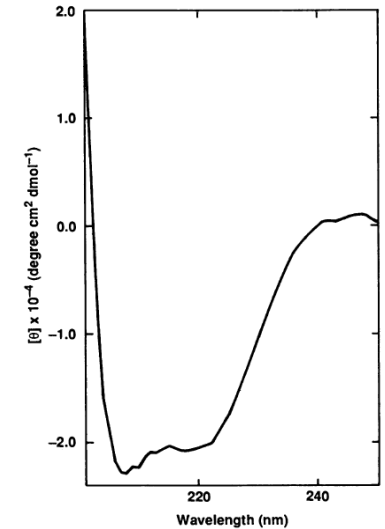
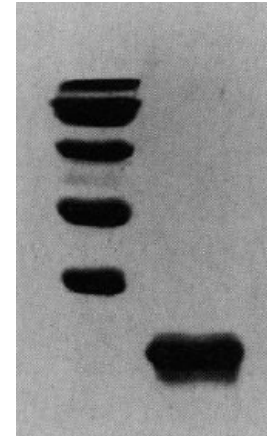


Design details

Helix: GELEELLKKLKELLKG, **Loop:** PRR

α_4 : Met-**Helix**-**loop**-**Helix**-**loop**-**Helix**-**loop**-**Helix**-COOH

Regan and DeGrado, Science 241, 976 (1988)



Binary patterned helix can lead to unpredicted oligomeric states—a 12 residue fragment accidentally synthesized during the study (ELLKKLLEELKG) self-associates into a mix of hexamer and tetramer

Hill et al, Science 249, 543 (1990)

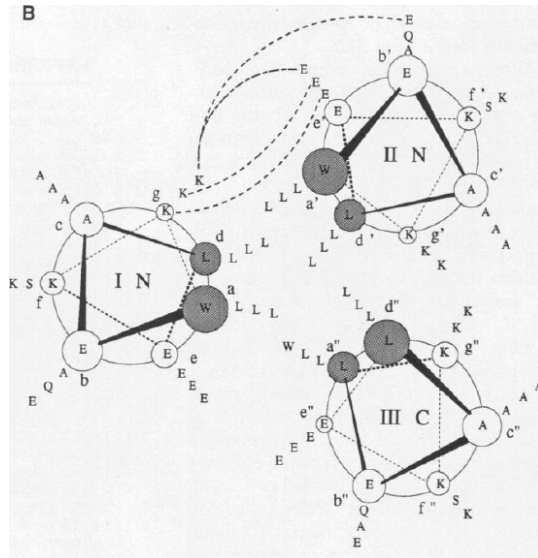
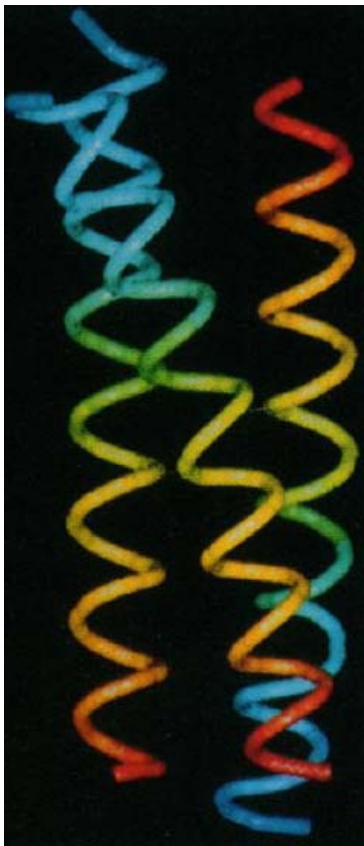
Protein	$\Delta G^{\text{H}_2\text{O}}$ (kcal mol ⁻¹)
α_4	-22.5
Lysozyme	-8.9
Myoglobin	-7.6
Ribonuclease A	-7.5
α -Lactalbumin	-4.2

Not all amphiphilic helices associate into a four helical bundle
Side chain interdigitation is necessary—computer modeling

Coiled-coil design

“Coil-Ser” was designed to form a coiled-coil based on previous studies with the helical heptad repeat in mind (i.e. “a” and “d” hydrophobic = Leu) studies show L is preferred at “d” whereas L, I, V, M are tolerated at “a”

Position	g	abcdefg	abcdefg	abcdefg	abcdefg	abcdef
Hodges	K	LEALEGK	LEALEGK	LEALEGK	LEALEGK	LEALEG
GCN4-p1	R	MKQLEDK	VEELLSK	NYHLENE	VARLKKL	VGER
coil-Ser	E	WEALEKK	LAALESK	LQALEKK	LEALEHG	



Why a 3 helix bundle rather than a coiled coil?

- neutralization of macrodipole moment
- intermolecular salt bridges
- more favorable side chain dihedral angles

Lovejoy et al, Science 259, 1288 (1993)

Determinants of coiled-coil topology

Coiled coil proteins with a heptad repeat sequence can form dimers, trimers and tetramers

Geometric properties of buried residues may influence the overall structure

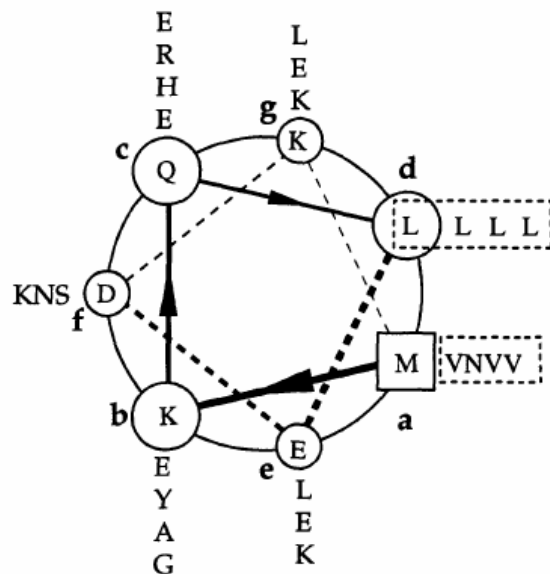


Table 1. Core mutants of GCN4-p1 form stable two-, three-, and four-helix structures.

Positions*		$-\left[\theta\right]_{222}$ (deg cm ² dmol ⁻¹)	T_m (°C)	$T_m^{\text{GdmCl}\dagger}$ (°C)	No. of helices‡	
a	d				Unmodified	SS
	GCN4-p1	33,300	53	<0	2	2
I	L	32,400	>100	77	2	2
I	I	32,400	>100	70	3	6
L	I	30,600	>100	94	4	4
V	I	22,500§	73	<0	—	(4,6)
L	V	30,600	81	<0	3	(2,—)
V	L	32,400	95	49	(2,3)	2
L	L	31,500	>100	76	3	(2,4,6,—)

*The residues inserted at four **a** and four **d** positions of GCN4-p1 (41). † T_m^{GdmCl} denotes the melting temperature in 3 M GdmCl. All scans and melts were performed at 10 μM peptide concentration. ‡The number of helices in the solution complex formed by unmodified peptides and by disulfide bonded peptides. The first four peptides were assigned on the basis of equilibrium analytical ultracentrifugation data and the last four on the basis of gel filtration data. Parentheses indicate that multiple species were present; a dash indicates the presence of a species that could not be assigned. §p-VI exhibits a $-\left[\theta\right]_{222}$ value of 31,500 deg cm² dmol⁻¹ at 150 μM concentration.

Designing new topology

All coiled-coil proteins are left-handed

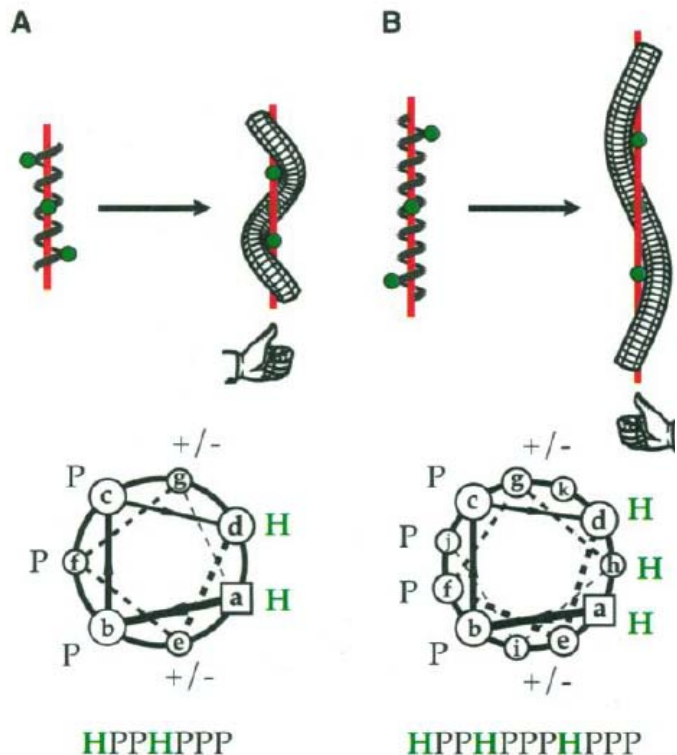
- 3.6 residues/turn \rightarrow 7 residues = $700^\circ < 2$ complete turns
- entire helix needs to coil around the superhelical axis by 20° every 7 residues to maintain hydrophobic contacts

Is it possible to design a right-handed coiled coil?

- a sequence based on an 11 residue repeat
- $1100^\circ > 3$ complete turns (by 20°)

Incidentally, test the effect of burying hydrophobic residues (a, d, h, or 1, 4, 8) on defining a topology

Harbury et al, Science 282, 1462 (1998)



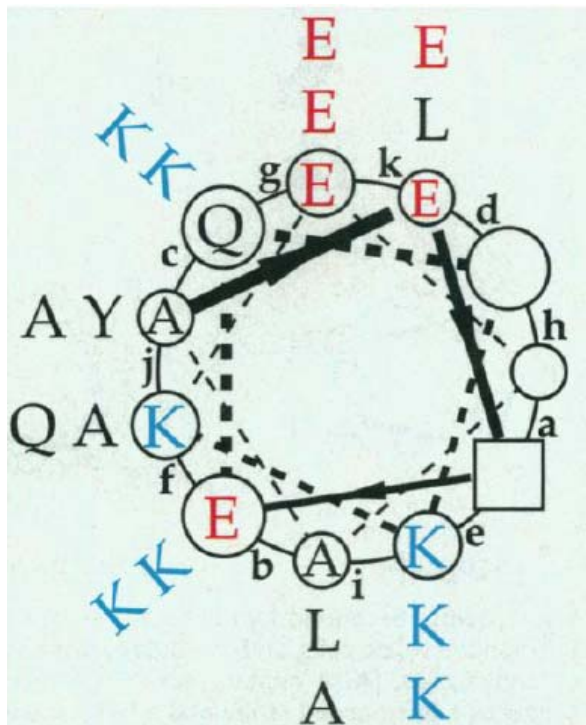
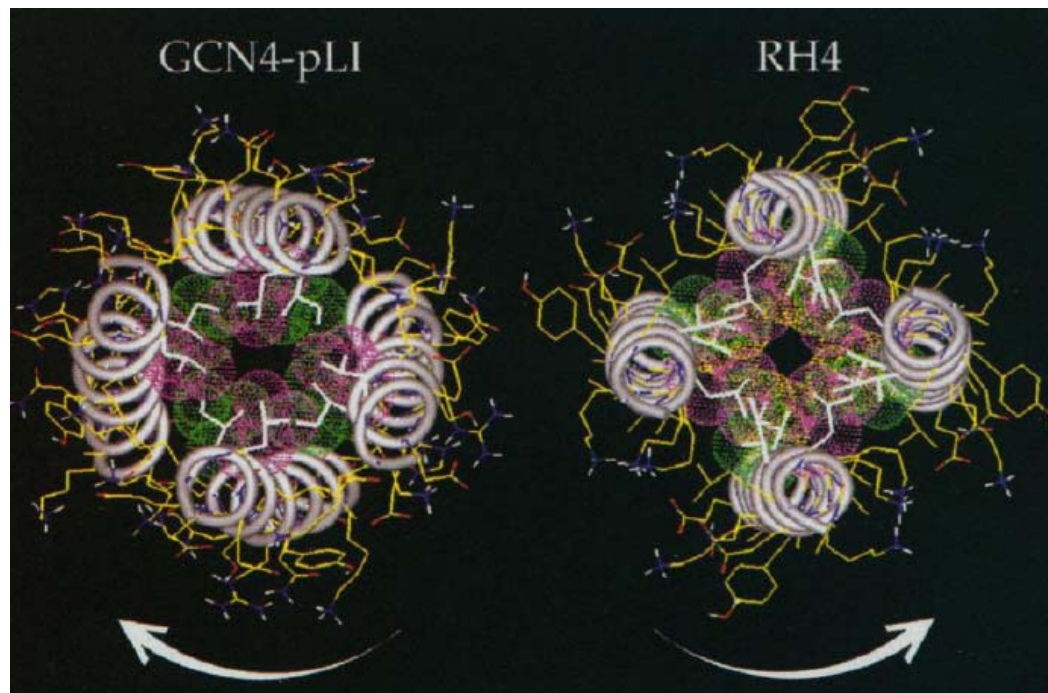
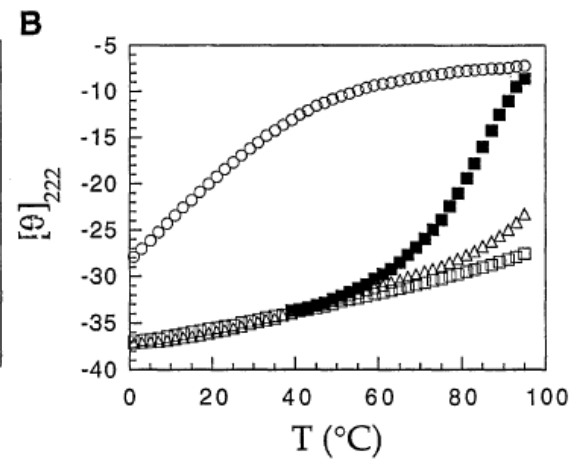
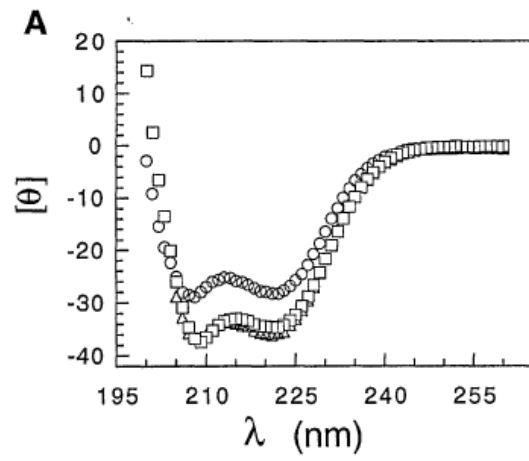


Table 1. Top sequence solutions for right-handed dimer, trimer, and tetramer coiled coils according to the packing calculation. The table consists of three lists, the first sorted according to dimer stability, the second according to trimer stability, and the third according to tetramer stability.

Sequence†			Stability‡	Specificity§	
a	d	h		Dimer-trimer	Dimer-tetramer
Dimer					
nV	L	nV	2.0	0.9	1.3
nV	nV	L	1.9	0.8	0.7
nV	L	L	1.9	0.7	0.9
l	L	nV*	1.8	1.8	1.0
Trimer				Trimer-dimer	Trimer-tetramer
nV	al	L	2.6	0.3	-0.5
l	al	L	2.4	0.5	0.2
l	al	nV	2.4	0.6	0.6
nV	al	nV	2.3	0.6	-0.4
L	al	L	2.0	-0.1	-0.8
al	al	l*	2.0	1.9	2.0
Tetramer				Tetramer-dimer	Tetramer-trimer
nV	al	l	3.1	2.2	1.7
nV	al	L	2.6	0.6	0.5
L	al	l*	2.5	1.9	2.0
L	al	L	2.3	0.3	0.8

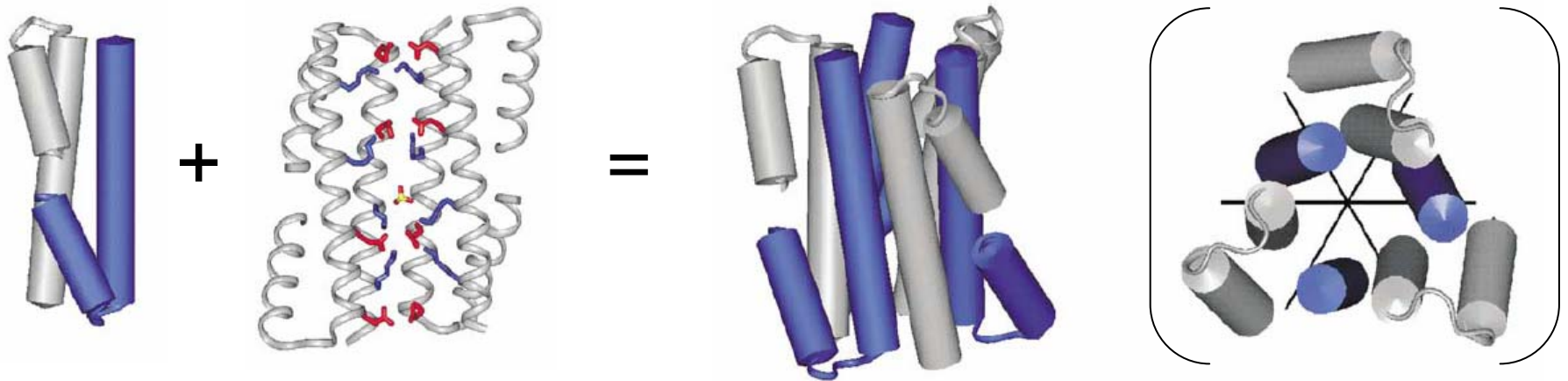
al = alloisoleucine, nV = norvaline



Hexameric helix bundle

Apply symmetry to simplify the design task and engineer a bigger protein complex

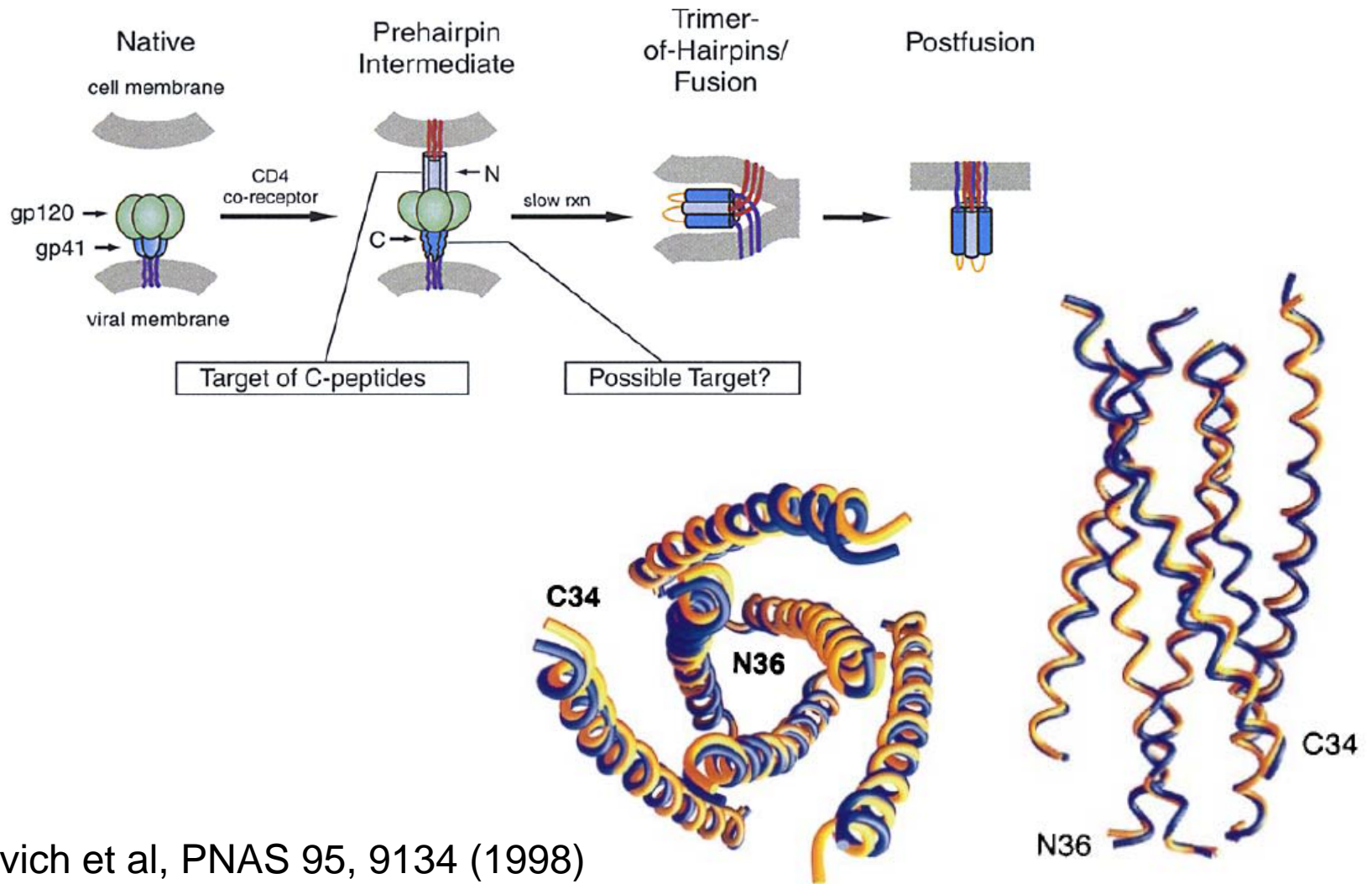
Introduce salt bridges at the interface to stabilize interaction



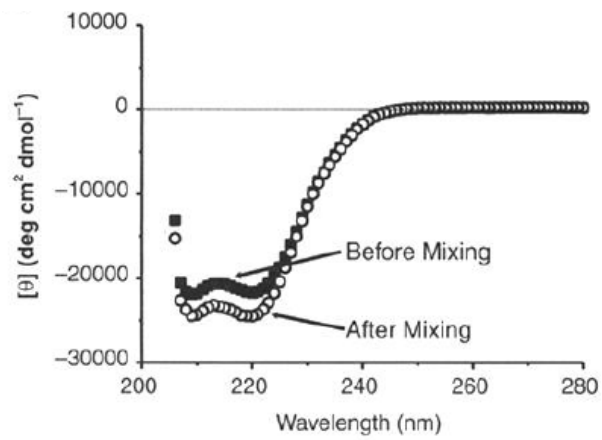
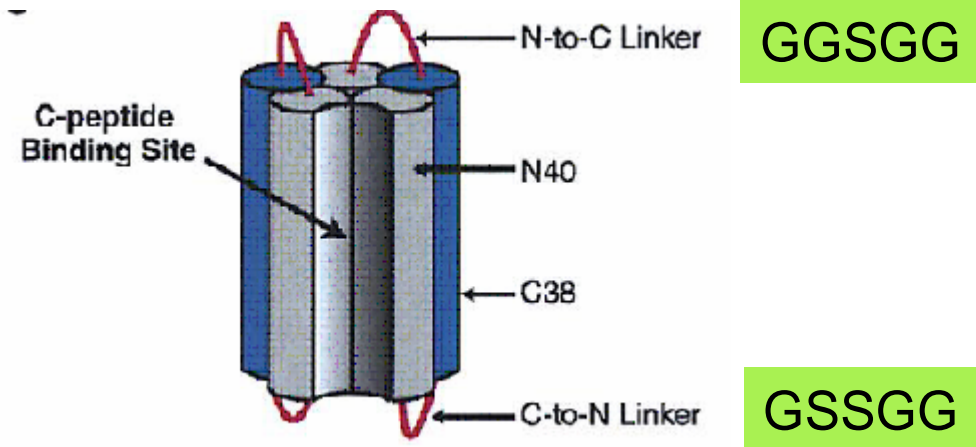
Ghirlanda et al, JMB 319, 243 (2002)

Helix bundle as HIV-1 inhibitor

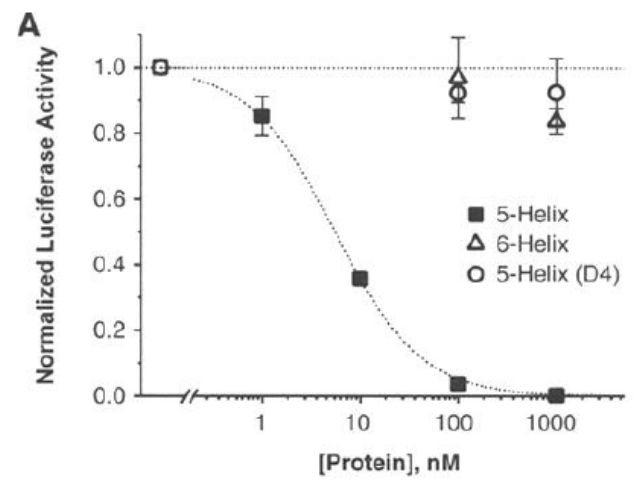
Helix bundle may be engineered for medical applications



Malashkevich et al, PNAS 95, 9134 (1998)



infectivity assay



Root et al, Science 291, 884 (2001)

Anticancer helix

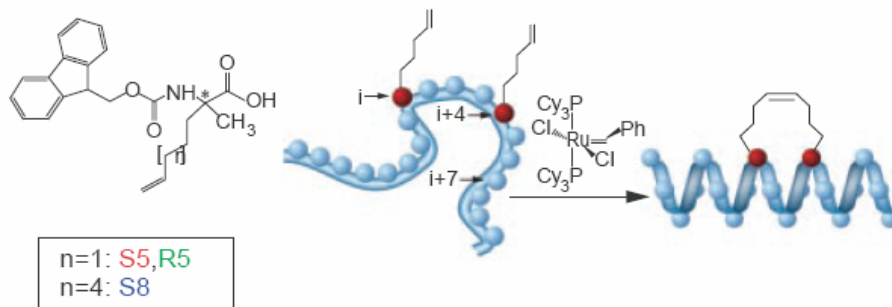
Bcl-2 regulates apoptosis in B-cells through conserved alpha helical domain (BH domain or “death” domain)

anti-apoptotic: Bcl-2, Bcl-X_L

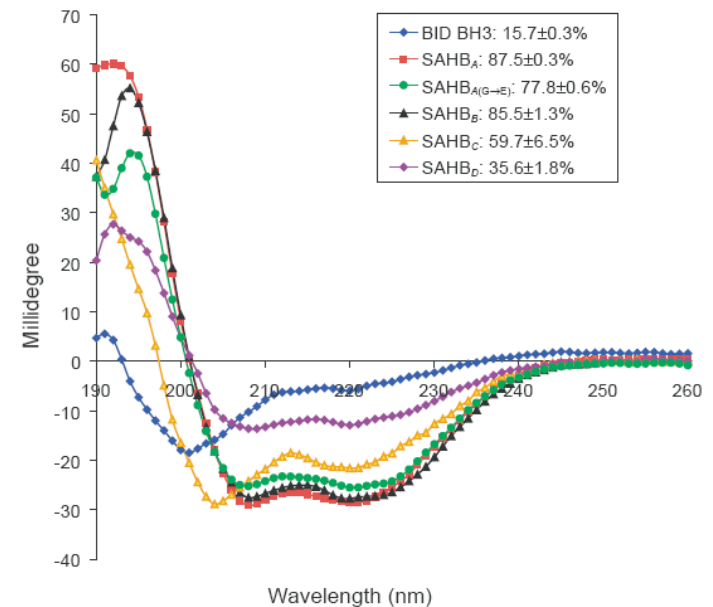
pro-apoptotic: BAX, BAK, **BID**, BAD

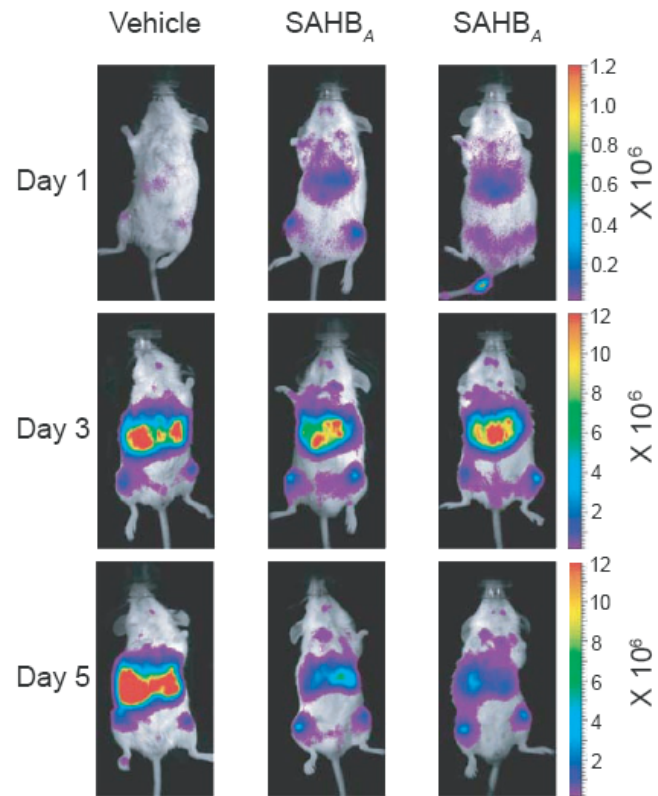
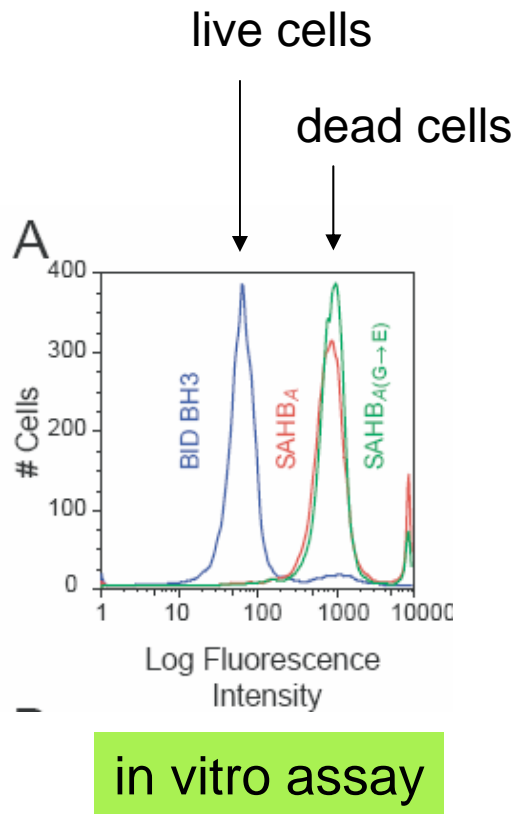
BH helix from BID can sequester anti-apoptotic Bcl-2 proteins, but is unstable as isolated helix

Hydrocarbon “staple” across the helix turn can stabilize the helix



Walensky et al, Science 305, 1466 (2004)





red = leukemic cells

Beta sheet design

The factors that modulate the formation and stability of beta sheet protein are not well understood

- both local and global effects are important
- stabilizing factors become apparent only in large structures
- long distance interactions within a beta sheet are difficult to engineer systematically
- contrasts with alpha helix

Practical concerns

- designed beta hairpins and beta sheets tend to have low solubility and easily aggregate—in part due to exposed main chain polar atoms
- amino acids of high beta sheet propensity are in general hydrophobic, e.g. Val, Ile, Phe, Tyr

Hierarchic design of beta sheet protein

Can an iterative hierarchic approach be applied to design a soluble monomeric beta sheet protein so that it folds without a metal or a disulfide bond?

- small proteins containing disulfide bonds are classified separately in SCOP

Information available that may be used during hierarchical design

- experimental data on beta hairpin stability
- amino acid beta sheet propensities
- statistical preferences for interstrand residue pairs
 - Wouters and Curmi, Proteins 22, 119 (1995)
- side chain rotamer modeling

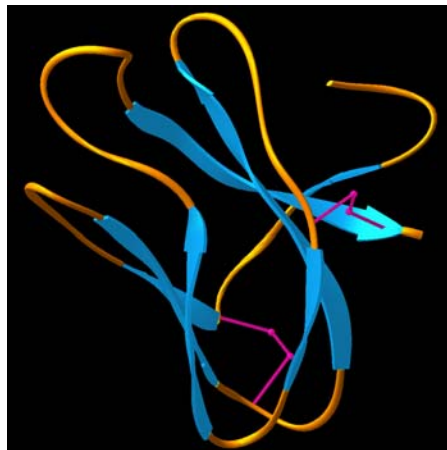
TABLE IV. High Scoring Pairs*

H-bonded		Non-H-bonded	
Cys-Cys	4.9	Cys-Cys	9.9
Glu-Lys	3.4	Glu-Lys	3.2
Glu-Arg	3.4	Asp-His	3.0
Gln-Arg	2.5	Ser-Asn	2.1
Phe-Phe	2.4	Thr-Thr	2.0
Ser-Ser	2.2		
Asp-Lys	2.1		
Gln-Lys	2.1		
Thr-Asn	2.0		

Beta hairpin peptide

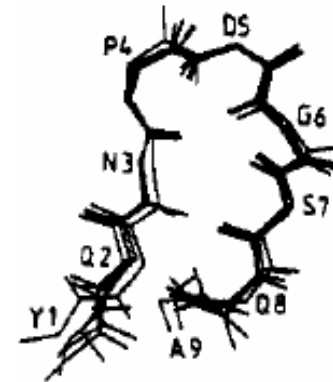
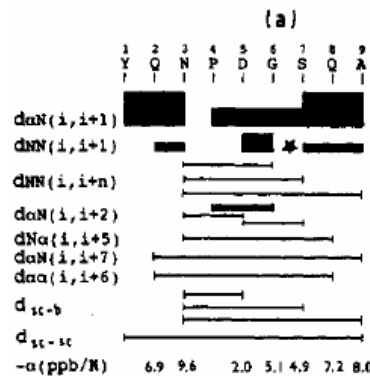
Beta turns form early during protein folding and stabilize chain reversal

Early designs of non helical structures included beta hairpins containing 8-16 residues



tendamistat

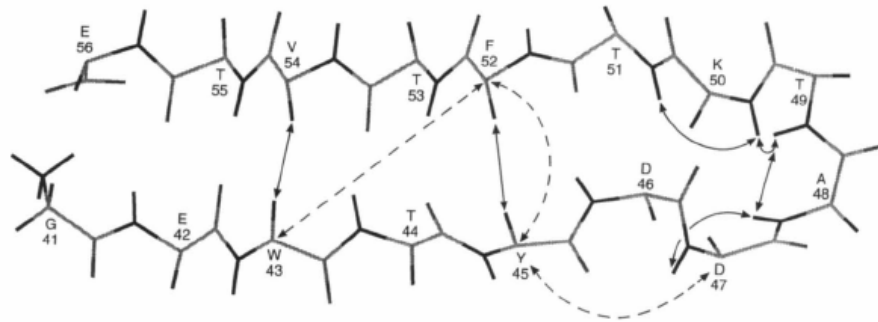
YQSWRYSQA (wt)
YQNPDGSQA (designed)
GHNPDGHG (neg control)



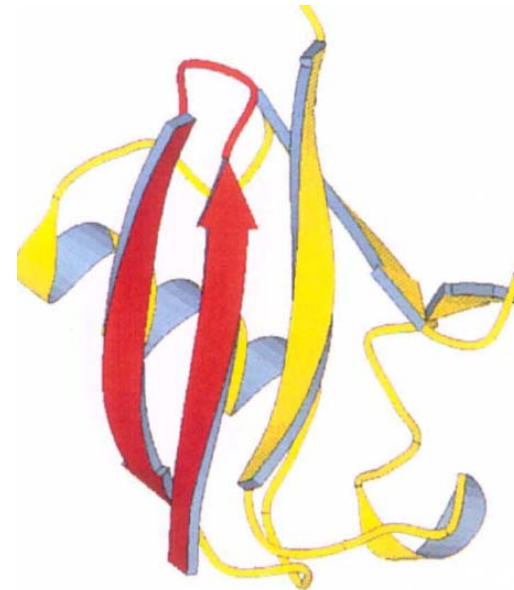
long distance NOE provide the most conclusive evidence

Blanco et al, JACS 115, 5887 (1993)

Some peptides isolated from natural proteins containing beta hairpin form beta hairpins in vitro, while others do not



Protein G, B1 domain 41-56
Blanco et al, NSB 1, 584 (1994)



ubiquitin residues 1-17 require optimization of the turn sequence
TLTGK → **NPDG**

Searle et al, NSB 2, 999 (1995)



But not the first hairpin of GB1
Blanco et al, Biochem 33, 6004 (1994)

Loop residue chirality

D-amino acids intrinsically prefer type I' and II' turns (why would this be?)

MQIFVKSXXKTITLVKV

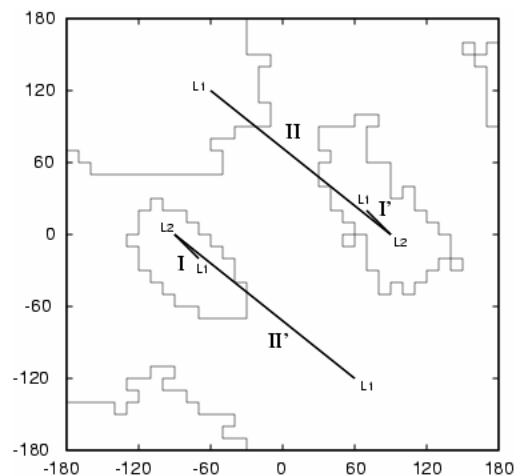
L-P-L-A (x)

D-P-D-A (✓)

D-P-L-A (✓)

D-P-G (✓)

L-P-G (x)



Haque & Gellman, JACS 119, 2303 (1997)

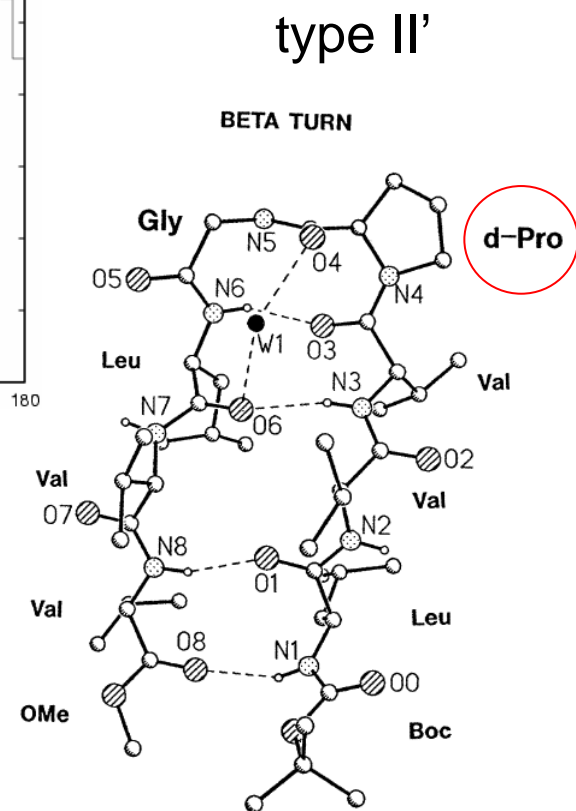
RYVEVXGOKILQ

D-P (✓✓) > L-N (✓)

L-P (x)

○ = ornithine

Stanger & Gellman, JACS 120, 4236 (1998)



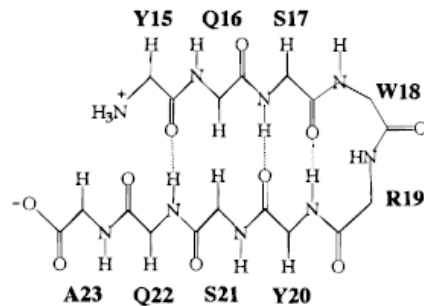
Karle et al, PNAS 93, 8189 (1996)

Turn residues are key

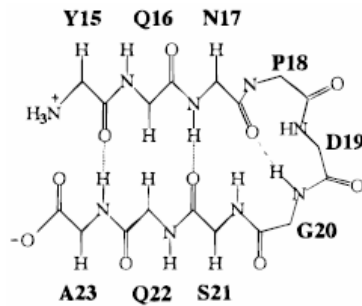
Turn conformation depends on the sequence of the loop

Interactions involving side chain of residues before and after the turn (inter-strand interactions) can further influence the turn stability and geometry

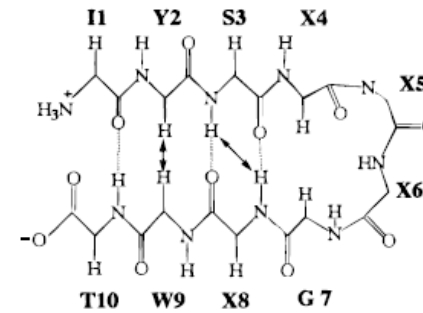
Peptide	X4	X5	X6	X8
2	N	P	D	G T
3	N	S	D	G T
4	N	S	D	G V
5	A	P	D	G T
6	A	K	A	G T



a) β -hairpin 2:2, type I β -turn



b) β -hairpin 3:5

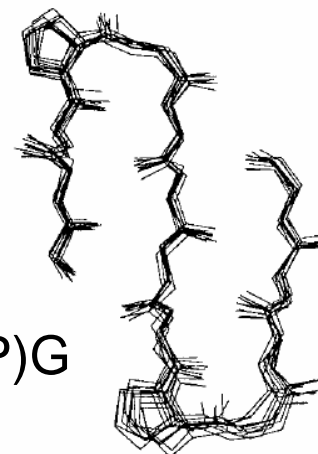


c) β -hairpin 4:4

Three stranded sheet

(Slightly longer) peptides with two turn sequences

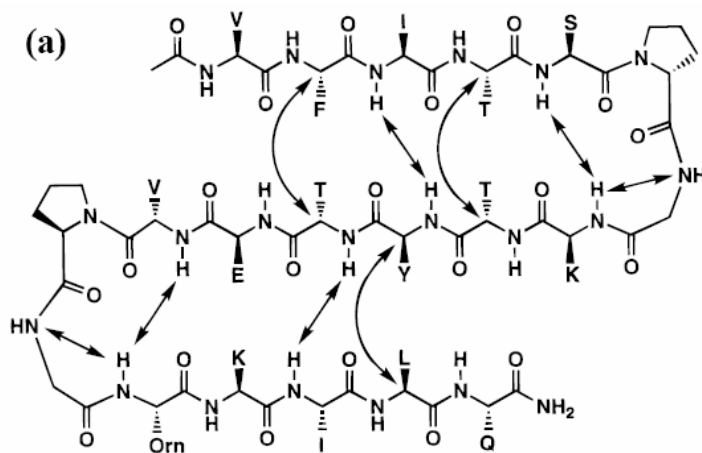
- optimization of folding and solubility (often requires non-aqueous solvent)
- not as easy as it seems
- works better with D-amino acids



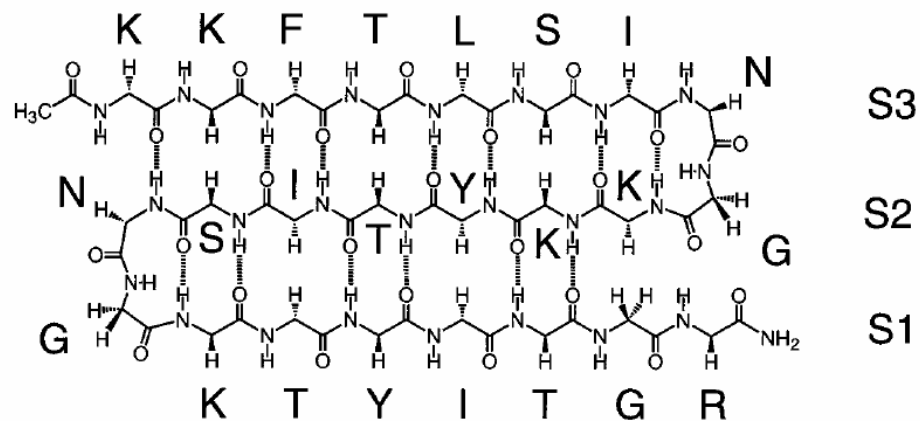
LFV(DP)GLVLA(DP)G
FVL

in chloroform/DMSO
Das et al, JACS 120, 5812 (1998)

Ac-VFITS(DP)GKTYTEV(DP)GOKILQ-NH₂



Schenck and Gellman, JACS 120, 4869 (1998)



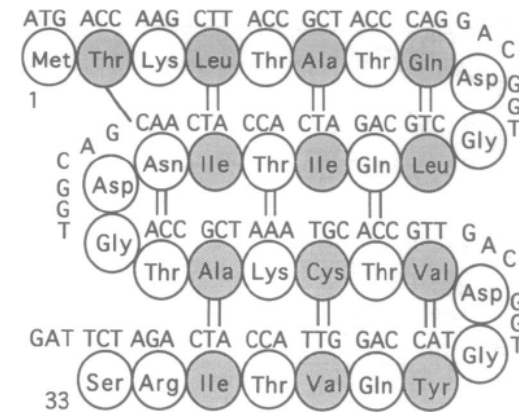
Aqueous methanol
Sharman and Searle, JACS 120,
5291 (1998)

Betadoublet

Beta sandwich topology is common in nature and include Ig, Con A, etc

Design and characterize a beta sandwich protein de novo

- Construct backbone using poly-ala ($\phi = -139^\circ$, $\psi = 139^\circ$), twist the sheet, and further modify ϕ/ψ angles of residues for interstrand hydrogen bond
- Type I' turns throughout—XDGX
- Sheets oriented at 30° of each other
- Disulfide bond between cys
- “Negative design” to disfavor Greek key—short turn sequence



Quinn et al, PNAS91, 8747 (1994)

Analysis by size exclusion chromatography

improved solubility (> 10 mg/ml), good dispersion of NMR peaks, cooperative thermal melting, reasonable CD, minimal ANS binding

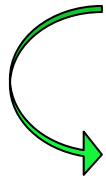
Also read about “Betabellin” by Yan & Erickson, Protein Sci 3, 1069 (1994)

Betanova

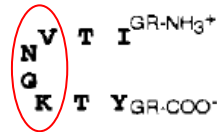
Start from a known hairpin structure and extend it long enough to constitute antiparallel beta strands

Ref. Griffiths-Jones, JMB

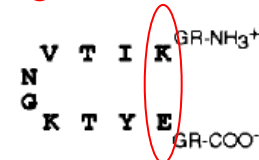
fold structure
35% to 45%



Hairpin I



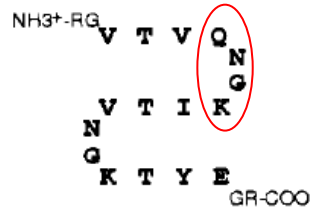
Hairpin II



NMR shows β -hairpin in equilibrium with random coil conformations. No cooperative unfolding behavior.

turn 1—ideal type I'

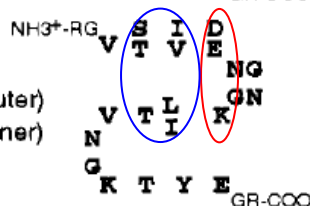
Sheet I



NMR shows β -hairpin formed by β -strands 2 and 3 in equilibrium with random coil conformations. No cooperative unfolding behavior.

- interstrand packing
- ionic pair

Sheet II (outer)



Sheet III (inner)

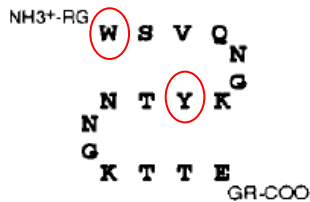
Addition of 40% TFE to Sheet I promotes formation of a three-stranded β -sheet.



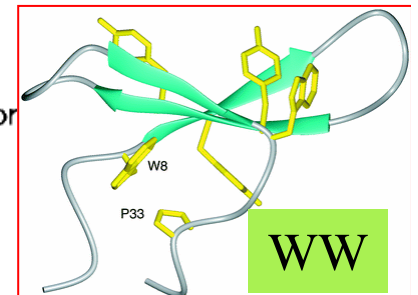
not much improvement

more hydrophobic residues in strands 1 and 2

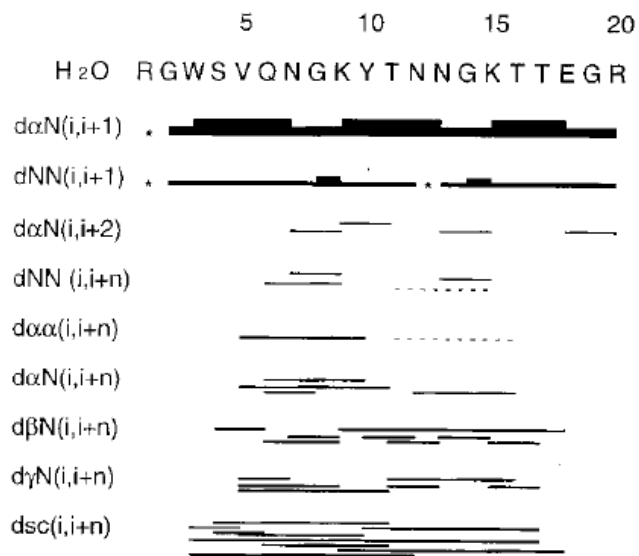
Betanova



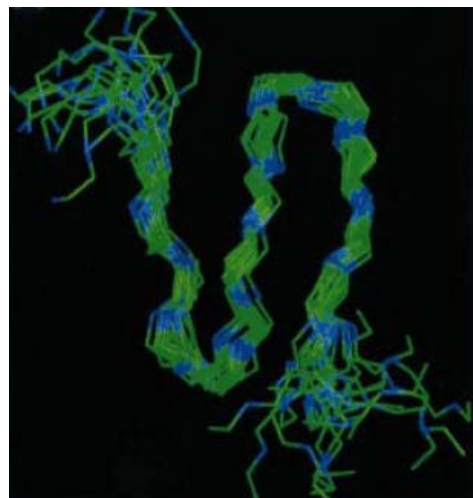
NMR shows a well defined three-stranded β -sheet. Cooperative unfolding behavior followed by fluorescence and CD spectroscopy.



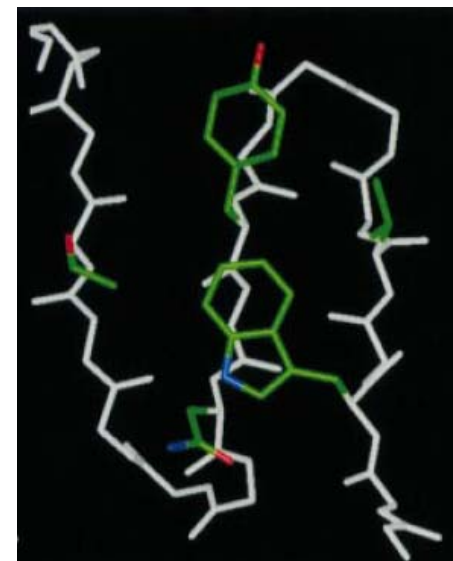
Kortemme et al, Science 281, 253 (1998)



NOE cross peak intensities



20 NMR structures

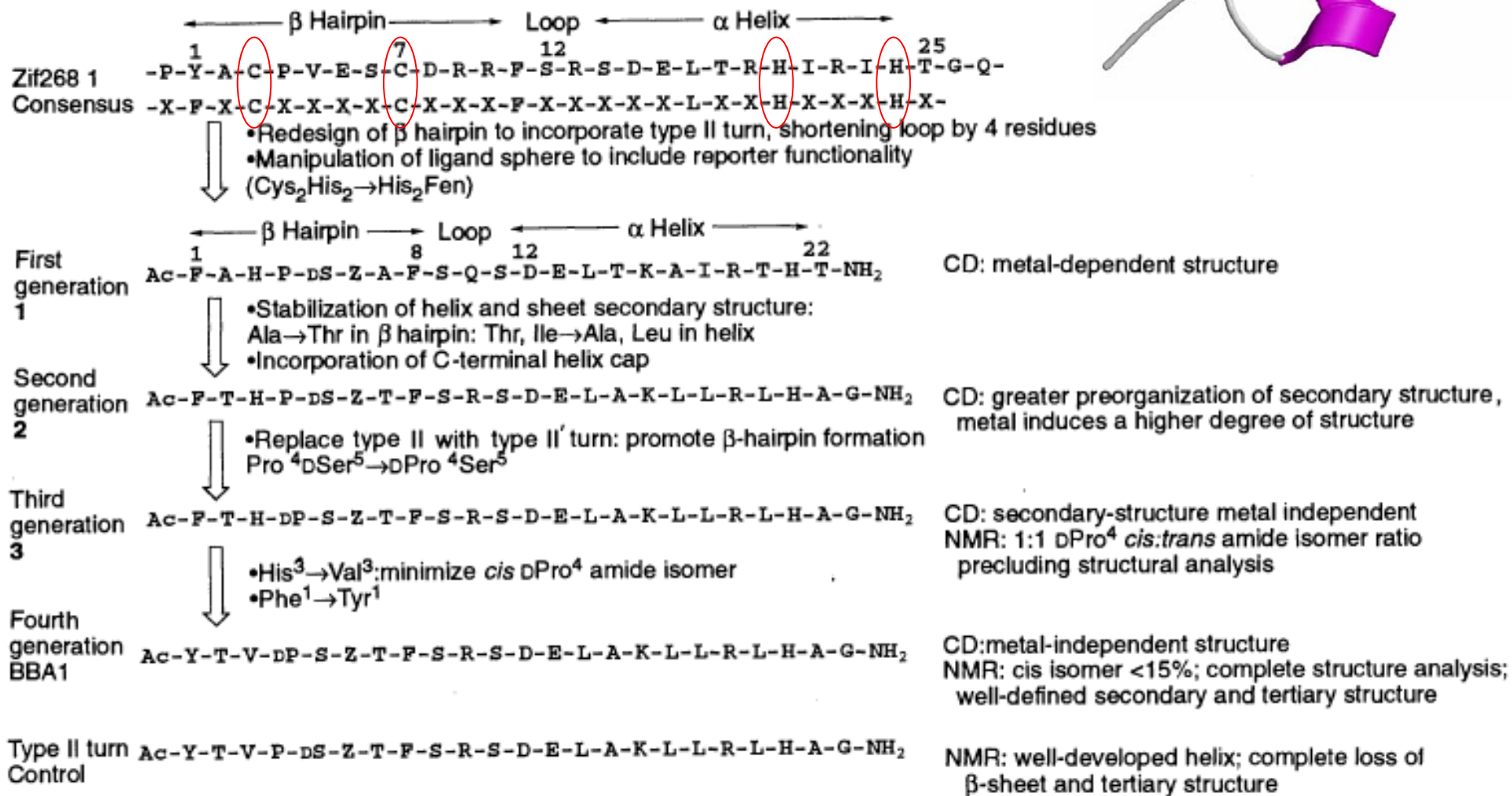
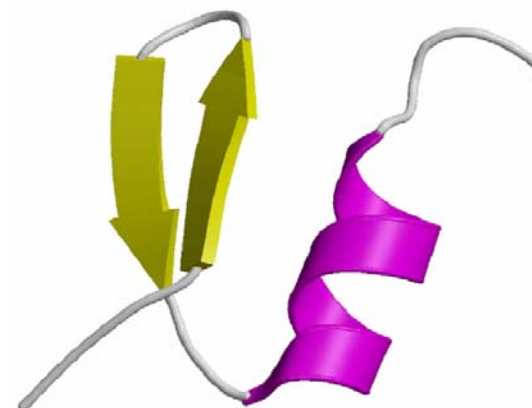


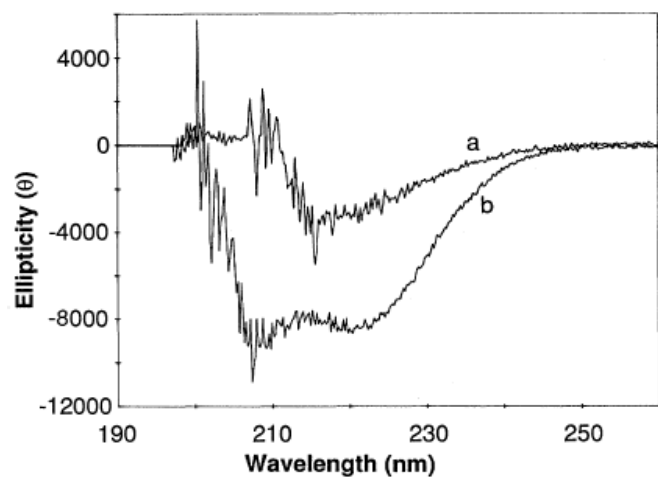
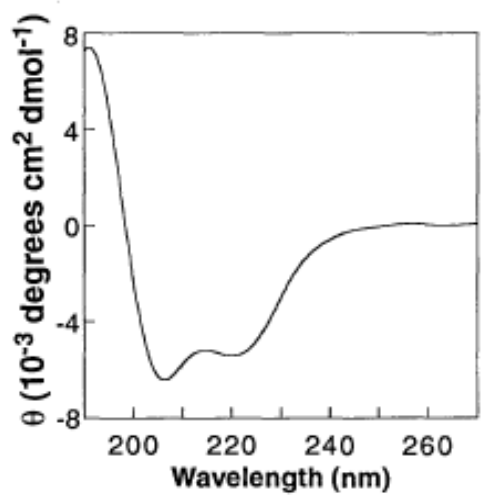
energy minimized structure

First example of a soluble three strand beta sheet protein made of natural amino acids stabilized purely through noncovalent bonds

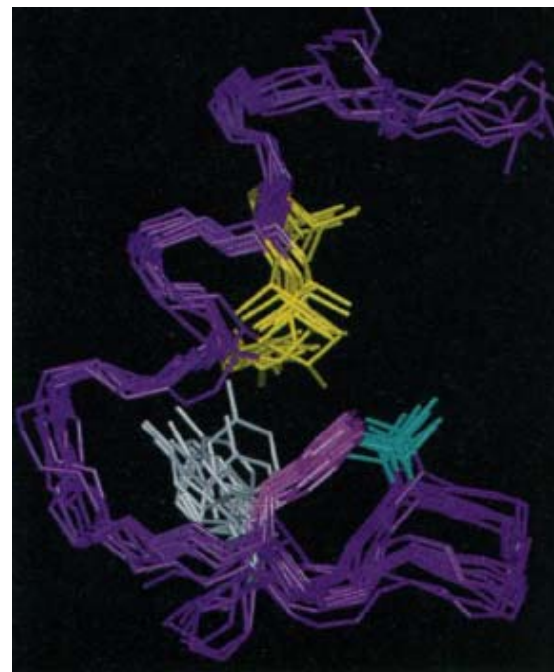
Protein of mixed topology

Design soluble small $\beta\beta\alpha$ motif domain

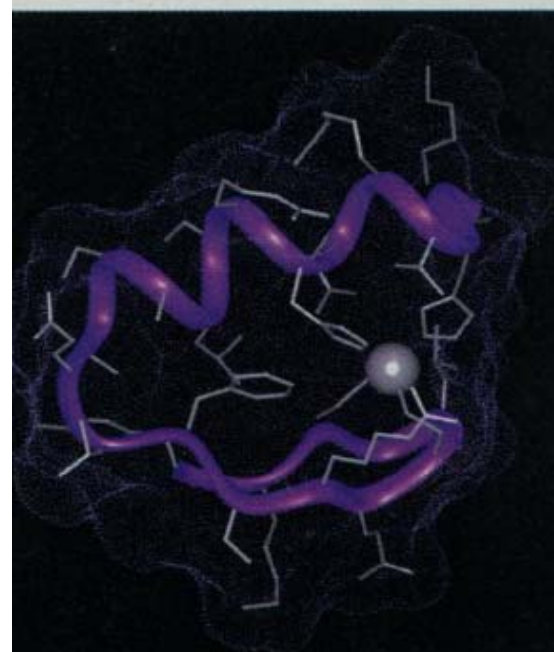




known zinc finger CD spectra



BBA1



Zif268

State of knowledge based design

Helices and simple helical proteins (coiled coils and helical bundles) can be reliably designed

Even simple designs may have profound biomedical implications (HIV inhibitor and cancer therapeutics)

Ability to design beta sheet proteins lags about 10 years

Intuition plays a critical role in all design problems